



Parental Polymorphism between Samba Mahsuri and False Smut Tolerant Landraces using SSR and InDel Markers

Preeti¹, Loksha R³, Balakrishnan D², BhaskarM², Gireesh C², Neeraja CN², Singh AK⁴, Nidagundi JM¹, Diwan JR¹, Bheemanna M¹, Suma TC¹, Prasad MS², Sundaram RM² and Ladhalakshmi D^{2*}

¹University of Agriculture Sciences, Raichur, Karnataka, India - 584104

²ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India - 500030

³University of Agricultural Sciences, Shimoga, Karnataka, India - 577201

⁴ICAR-Indian Agricultural Research Institute, New Delhi, India - 110012

*Corresponding author Email: ladhasavitha@gmail.com

Received : 9th April, 2023; Accepted : 6th June, 2023

Abstract

In the present study, 868 SSR and 475 InDel markers were selected to study parental polymorphism of two different crosses between recipient parent Samba Mahsuri and false smut donor lines IC379047 and IC334233 at ICAR-Indian Institute of Rice Research, Rajendranagar, Hyderabad. Out of 1343 markers used for parental polymorphism, 100 markers were polymorphic between parental lines Samba Mahsuri and landrace IC379047 and the total polymorphism percentage recorded was 7.44%. Ninety-nine markers exhibited polymorphism between parental lines Samba Mahsuri and IC334233 and 7.37% of total polymorphism percentage was recorded between the parents. The identified polymorphic markers could be further used for QTL mapping studies in false smut resistance.

Keywords: Rice, false smut, polymorphism, SSRs and InDel markers.

Introduction

Rice (*Oryza sativa* L) is a widely cultivated cereal crop all over the world. It is a nutritionally indispensable food crop with carbohydrates being the major fraction along with protein and vitamins; and 21% of energy for more than half of the world's population. The major rice producing countries are China which ranks first in production of 148.3 million metric tons followed by India and Indonesia with production of 122.27 and 35.3 million metric tons respectively (UASD, 2021). To meet the demand of increasing human population, the higher rice productivity can be achieved by developing varieties having stable yielding ability across the areas along with resistance to various pest and diseases. The germplasm consisting of landraces, modern cultivars, breeding stocks, wild

forms and wild relatives of the cultivated crop species could be the valuable genetic source to identify the promising donors. Germplasm refers to sum total of genetic material *i.e.*, possible alleles of the various genes present in crop species and its wild relatives, which can be used for exploiting the genes governing various traits for biotic and abiotic stresses; and ultimately in breeding of the new variety. In previous studies at ICAR-IIRR, Hyderabad, landrace IC379047 (Mancha) and germplasm IC334233 were identified as a tolerant source for the false smut disease through artificial screening. The identified lines were crossed to the popular high yielding but disease susceptible cultivar Samba Mahsuri (an elite fine-grain *indica* rice cultivar). The two parental genotypes in crosses

viz., Samba Mahsuri x IC379047 and Samba Mahsuri x IC334233 were phenotypically diverse and genomic diversity was assessed using the marker system.

Molecular markers have wide applications in several genetic research and breeding programme such as genetic diversity assessment, Quantitative Trait Loci (QTL) identification, gene mapping, marker-based gene targeting and characterization of alien introgression lines from wild species of rice. DNA markers like Restriction Fragment Length Polymorphism (RFLP), Random Amplified Polymorphit DNA (RAPD), Sequenced Tagged Sites (STS), Cleaved Amplified Polymorphic Sequence (CAPS) and Simple Sequence Repeats (SSR) have been used in molecular studies over the past few decades. PCR and gel-based markers have several practical utilities for the researchers and breeders. In addition, the recently identified InDel markers are user friendly and also has ease of accessibility and technical simplicity. These markers have been attained great importance over the last two decades in several genotyping studies because of their co-dominant nature, reproducible, high polymorphism, multiallelic nature, widely distributed across the genome, require less DNA quantity and are cost effective (Usman *et al.*, 2018 and Chukwu *et al.*, 2019). The SSR markers are present in both coding and non-coding genomic region with lower level of mutation rate (10^{-2} and 10^{-4}) per generation. They are used in several studies such as population structure and evolutionary studies, linkage map construction, genetic mapping and marker assisted selection (Edwards and Balley, 2010 and Gonzaga *et al.*, 2015). QTL mapping requires mapping population and sufficient number of polymorphic markers identified between the parental lines. Different mapping populations are used for mapping studies like double haploids, F_2 populations, F_2 derived F_3 population, near isogenic lines (NILs), and recombinant inbred lines (RILs).

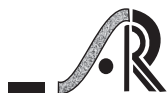
Each mapping population has its own advantages and disadvantages. The polymorphic markers identified between the parental lines are used for the genotyping of mapping population. Phenotypic and genotypic data of mapping population was used for the QTL mapping using mapping tools. The main objective of our research was to identify the polymorphic SSR markers between the parental genotypes for further QTL mapping studies for false smut disease tolerance.

Materials and Methods

The experimental material for the study comprised of false smut donor lines IC379047 and IC334233 and recipient genotype Samba Mahsuri (high yielding variety susceptible to false smut disease), and SSR and InDel markers were collected from the Department of Genetics and Plant Breeding, ICAR-IIRR, Hyderabad.

Young leaf samples were collected from the field and plant genomic DNA was isolated by CTAB method (Doyle and Doyle, 1987). The leaf samples were ground in CTAB buffer using pestle and mortar. The ground samples were transferred into Eppendorf tube and incubated at 65 °C for 30 min. The samples were centrifuged for 15 min at 13000 rpm, the supernatant (genomic DNA) was transferred to an Eppendorf tube and equal amount of 24:1 chloroform and isoamyl alcohol was added and again centrifuged for 10 min at 13000 rpm. The supernatant was taken into new Eppendorf tube and equal amount of chilled isopropanol was added and incubated at -20 °C for 15 min. Centrifugation was done for 10 min at 13000 rpm. The pellet formed in the tube was washed with 70% chilled ethanol and kept out overnight to dry and stored at 4 °C in Tris EDTA (TE) buffer for further genomic study.

A total of 1343 SSR markers distributed on 12 rice chromosomes were used for the parental polymorphism survey. Among 1343 primers, 868 markers were selected from www.gramene.org. and



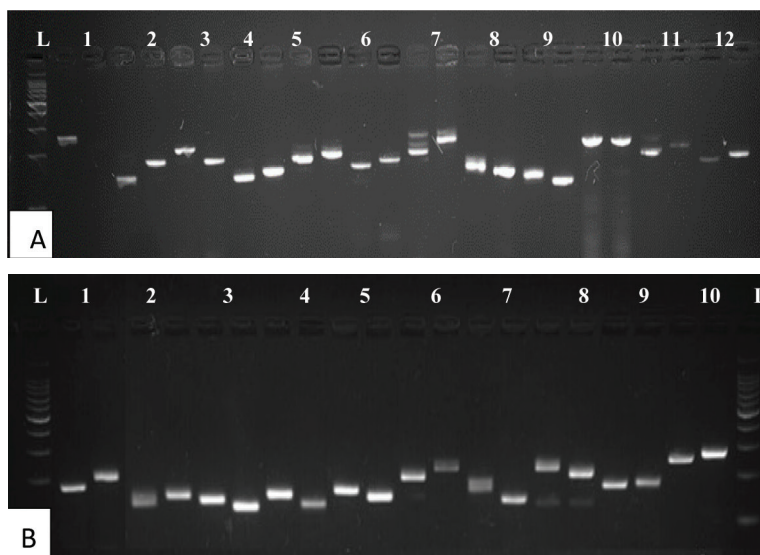
remaining 475 were newly developed InDel marker set chosen from the study conducted by Hechanova *et al.*, 2021. For PCR analysis, a total 10 µl PCR reaction mixture contained 30-50 ng of 2 µl genomic DNA template, 1 µl of 10X buffer, 5.2 µl of sterilized distilled water, 0.5 µl of 2.5 mM dNTP, 3 units of 0.3 µl Taq polymerase and 1 µl of primer was prepared and PCR amplification was carried out in Bio-Rad PCR machine. The thermal cycles were programmed as follows, mixture was incubated at 94 °C for 5 min; then 35 cycles of 1 min of denaturation at 94 °C, 30 sec of annealing at 58 °C and 1 min of extension at 72 °C; and 10 min of final extension at 72 °C. The PCR products were separated in 3.5% agarose gel with 1X Tris-borate-EDTA (TBE) buffer and the band sizes of the PCR products were detected and visualized through gel documentation unit. Graphical mapping of the markers on all 12 rice chromosome was done using web based tool *Oryza*BASE.

Results and Discussion

Landraces are reported as genetic reservoirs of many useful genes, which could be introgressed into the cultivars. With respect to rice false smut disease, resistance source has not been identified till date and the disease has been severely affecting the rice production and can become the great cause for the huge yield loss in future. Hence, there is a need to identify the resistance source against false smut. An attempt is made to identify the tolerant sources against false smut through artificial screening at ICAR-IIRR farm, Hyderabad. The tolerant sources for false smut disease were used as donors and high yielding but false smut susceptible variety Samba Mahsuri was used as the recipient parent. The polymorphism survey between two parental genotypes in two separate crosses *viz.*, Samba Mahsuri x IC379047 and Samba Mahsuri x IC334233 was carried out using 1343 SSR markers. The per cent of polymorphism was calculated by

formula *i.e.*, the number of polymorphic markers to the total number of markers for each chromosome multiplied by 100. Among the 1343 markers used in polymorphic study for the Samba Mahsuri x IC379047, 100 SSR markers were found to be polymorphic (**Table 1**). The total polymorphism percentage between parents Samba Mahsuri and landrace IC379047 recorded was 7.44%. The highest percentage of polymorphism was observed on chromosome 1 (14.08%) followed by chromosome 2 and 7 (10.71%) and lowest polymorphic percentage value was recorded on chromosome 12 (1.26%).

Among the total 1343 markers used for the polymorphism study for cross Samba Mahsuri x IC334233, and out of 868 rice markers chosen from Gramene, only 27 SSRs were found as polymorphic and were distributed on all rice chromosomes except on chromosome 7. Among 475 SSRs chosen from InDel marker set designed, 72 markers were found to be polymorphic and were also distributed among all the 12 rice chromosomes. It was found 99 out of 1343 markers as polymorphic between the two genotypes in the study (**Table 1**). The total polymorphism percentage between parents Samba Mahsuri and germplasm IC334233 recorded was 7.37%. The highest percentage of polymorphism was observed on chromosome 6 (10%), which means out of 142 markers positioned on chromosome 6, 13 markers were found to be polymorphic followed by chromosome 1 (9.15%) and lowest polymorphic percentage value was recorded on chromosome 9 (4.30%). Polymorphism percentage for all the 12 rice chromosome for two crosses are represented in **Table 2**. The banding pattern of the polymorphic markers for two crosses has shown in the gel image (**Figure 1**). The frequency distribution and representation of the polymorphic markers on rice chromosomes for two crosses has shown in **Figure 2** and **Figure 3**.



S. No	Marker	S. No	Marker
1	A09P06588	L	Ladder
2	A09P12377	1	RM23946
3	A09P21225	2	RM216
4	A10P08920	3	RM25031
5	A10P09104	4	RM21749
6	RM6470	5	RM28766
7	RM5543	6	RM5479
8	RM17624	7	RM17377
9	RM1204	8	RM18182
10	RM142	9	RM15981
11	RM3394	10	RM22763
12	RM427		

Figure 1: Identified polymorphic SSR and InDel markers between A. Recipient parent (Samba Mahsuri) and donor (IC379047); B. Recipient parent (Samba Mahsuri) and donor (IC334233)

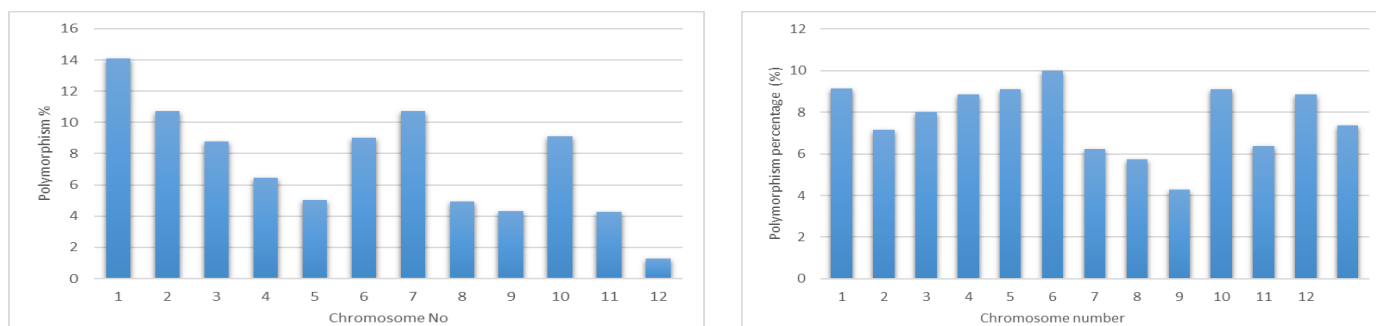


Figure 2: A. Frequency distribution of polymorphic markers identified between Samba Mahsuri x IC379047; B. Frequency distribution of polymorphic markers identified between Samba Mahsuri x IC334233

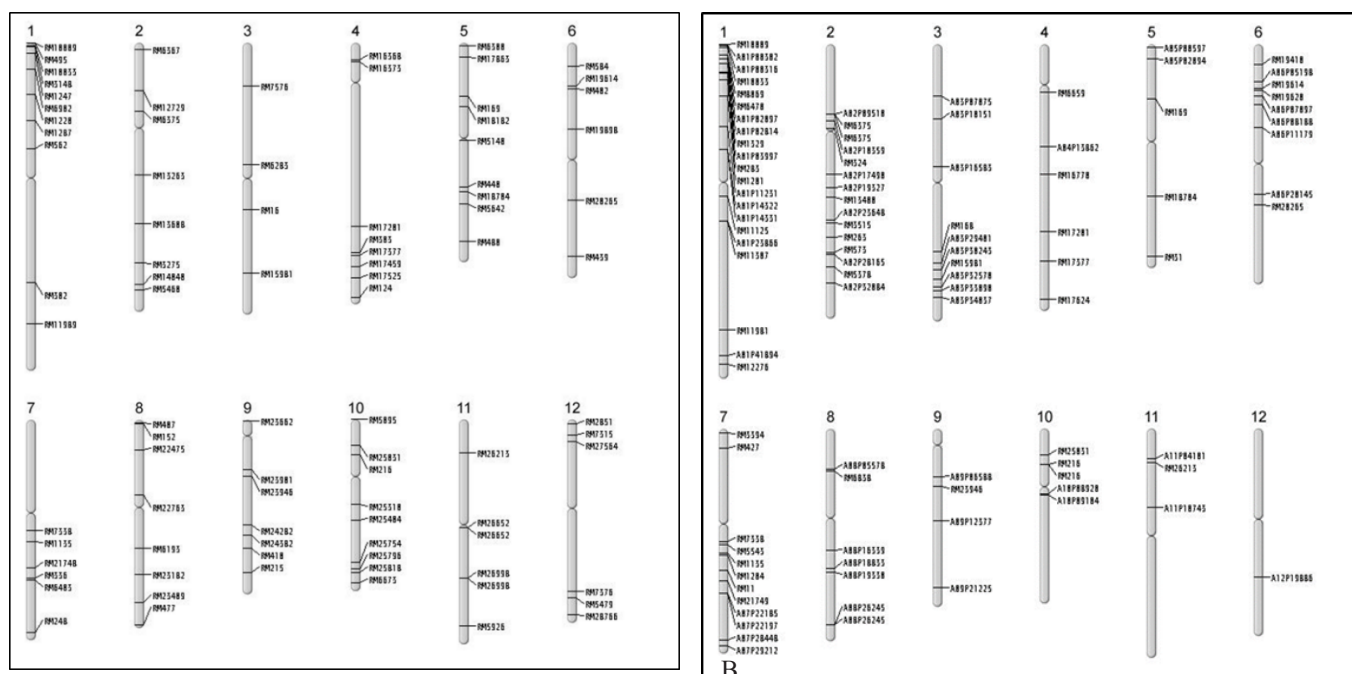


Figure 3: A. The distribution of the polymorphic markers on 12 rice chromosome for cross Samba Mahsuri x IC379047; B. The distribution of the polymorphic markers on 12 rice chromosome for cross Samba Mahsuri x IC334233

It was observed that the InDel marker set showed higher base pair difference between the parents compared to the SSR markers used. The polymorphic markers identified from the newly designed InDel marker set showed three times higher polymorphism than RM markers indicating that the InDel marker set exhibited higher allelic diversity between the parental lines. It was also reported that around 27 SSRs were found to be common in both the crosses across the genome except at chromosome 4,7 and 12 (Table 3). In addition to the parental lines, the

polymorphic markers identified between parents were used for the hybrid confirmation study, in which the markers clearly exhibited both the alleles in F_1 s indicating the true hybrid.

Similar studies on parental polymorphism have also been carried out by various researchers. Hable *et al.*, (2020) found the highest polymorphism percentage of 40.96% for chromosome 4 and on chromosome 9 recorded lowest polymorphic percentage (9%). Polymorphism percentage between the parents

Table 2: Polymorphism percentage for all the 12 rice chromosome for cross Samba Mahsuri x IC379047 and Samba Mahsuri x IC334233

S. No.	Chromosome No.	Total No. of markers used		No. of polymorphic markers obtained		Polymorphism (%)	
		Samba Mahsuri x IC379047	Samba Mahsuri x IC334233	Samba Mahsuri x IC379047	Samba Mahsuri x IC334233	Samba Mahsuri x IC379047	Samba Mahsuri x IC334233
1	1	142	142	20	13	14.08	9.15
2	2	140	140	15	10	10.71	7.14
3	3	125	125	11	10	8.80	8.00
4	4	124	124	8	11	6.45	8.87
5	5	99	99	5	9	5.05	9.09
6	6	100	100	9	10	9.00	10.00
7	7	112	112	12	7	10.71	6.25
8	8	122	122	6	7	4.91	5.74
9	9	93	93	4	4	4.30	4.30
10	10	55	55	5	5	9.09	9.09
11	11	94	94	4	6	4.25	6.38
12	12	79	79	1	7	1.26	8.86
Total markers		1343	1343	100	99	7.44	7.37

Table 3: Common polymorphic SSR and InDel markers identified between Samba Mahsuri x IC379047 and Samba Mahsuri x IC334233

Sl. No.	Markers	Chromosome No.	Sl. No	Markers	Chromosome No.
1	A01P00302	1	15	RM169	5
2	A01P11231	1	16	A06P05198	6
3	A01P23866	1	17	A06P08188	6
4	A01P02097	1	18	A06P11179	6
5	A01P37714	1	19	A06P20145	6
6	A02P10359	2	20	A08P05578	8
7	A02P17490	2	21	A08P16339	8
8	A02P09510	2	22	RM23946	9
9	A03P16583	3	23	A10P09104	10
10	A03P30243	3	24	RM216	10
11	A03P33090	3	25	RM25031	10
12	A03P10151	3	26	A11P10743	11
13	RM15981	3	27	A11P04101	11
14	A05P00597	5			



Rajendrakasturi and URG-30 reported was 29.02%. Chandu *et al.*, (2020) using 800 SSR markers observed 20.75% of total polymorphism percentage between Samba Mahsuri and *O. rufipogon* WR119 parents. The highest polymorphism percentage for chromosome 6 (26.67%) and lowest for chromosome 10 (8.93%) was also reported by Rathi *et al.*, (2021) with total percentage of polymorphism (16.67%) between Improved Samba Mahsuri and local landrace Badshabhog using 576 random SSR markers. Kulkarni *et al.*, 2020 found total polymorphism percentage of 6.93 between the parents IR58025A and KMR-3R using 1904 genomic SSR markers.

Conclusion

The identified polymorphic markers on all 12 rice chromosomes are useful for linkage analysis and QTL mapping for the traits of interest in the biparental mapping populations derived from the two crosses *viz.*, Samba Mahsuri x IC379047 and Samba Mahsuri x IC334233. Genotyping and QTL mapping using these markers for false smut resistance will help in mapping QTLs for resistance to false smut disease. Development of the false smut resistant rice varieties through marker assisted breeding method is feasible using the detected polymorphic markers flanking the QTLs.

Acknowledgment

The study was carried out as part of Ph.D. research work entitled “Genetic and molecular analysis of false smut resistance in landraces of rice (*Oryza sativa* L.)” of the first author at College of Agriculture, Raichur and ICAR-Indian Institute of Rice Research, Hyderabad, India. The work was supported under the SERB project on “Characterization and understanding the genetics of resistance of *Ustilaginoidea virens* and identification of false smut tolerant sources in rice” and DBT project on “Mainstreaming rice landraces diversity in varietal development through genome

wide association studies: A model for large-scale utilization of gene bank collections of rice”.

References

- Chandu G, Addanki KR, Balakrishnan D, Mangrauthia SK, Sudhakar P, Satya AK and Neelamraju S. 2020. SSR markers for grain iron zinc and yield-related traits polymorphic between Samba Mahsuri (BPT5204) and a wild rice *Oryza rufipogon*. *Electronic Journal of Plant Breeding*, 11: 841-7.
- Chukwu SC, Mohd Y, Rafii SI, Ramlee, Ismail SI, Oladosu Y, Okporie E, Onyishi G, Utobo E, Ekwu L, Swaray S and Jalloh M. 2019. Marker-assisted selection and gene pyramiding for resistance to bacterial leaf blight disease of rice (*Oryza sativa* L.), *Biotechnology and Biotechnological Equipment*, 33: 440-455.
- Doyle JJ and Doyle JL. 1987. A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochemical Bulletin*, 19: 11-15.
- Edwards D, Batley J. 2010. Plant genome sequencing: Applications for crop improvement. *Plant Biotechnology Journal*, 8:2-9.
- Gonzaga ZJ, Aslam K, Septiningsih E M, and Collard B C. 2015. Evaluation of SSR and SNP markers for molecular breeding in rice. *Plant Breeding and Biotechnology*, 3:139-152.
- Gramene [Internet]: a comparative resource for plants. 2019. Available from www.gramene.org.
- Hable S, Singh SK, Mounika K, Khaire A, Singh DK, and Majhi PK. 2020. Study of allelic variation at genome wide SSR loci in parents of mapping population for high grain zinc in rice (*Oryza sativa* L.). *Journal of Experimental Biological Agricultural Sciences*, 8: 558 - 575.
- Hechanova S, Bhattarai K, ElizaVie S, Graciana C, Pathmasiri K, Eok-KeunAhn, Li C, Lee J, Kohli AN, Hamilton RS, Hernandez JE, Gregorio GB,



- Kshirod KJ, Gynheung, and Kim SR. 2021. Development of a genome-wide InDel marker set for allele discrimination between rice (*Oryza sativa*) and the other seven AA-genome *Oryza* species. *Scientific Reports*, 11: 1-11.
- Kulkarni SR, Balachandran SM, Ulaganathan K, Balakrishnan D, Praveen M, Prasad AH, Fiyaz RA, Senguttuvel P, Sinha, P, Kale RR and Rekha G. 2020. Molecular mapping of QTLs for yield related traits in recombinant inbred line (RIL) population derived from the popular rice hybrid KRH-2 and their validation through SNP genotyping. *Scientific Reports*, 10: 1 - 21.
- Rathi S, Upadhyay S, Singh PK, Kumar R, Pallavi, Bisen P, Loitongbam B, and Sanchika S. 2021. Study of parental polymorphism and allelic variation for grain quality and yield traits in rice (*Oryza sativa* L.) Using SSR Markers. *International Journal of Plant Soil Science*, 33: 136-149.
- UASD, 2021. [https : // ipad.fas.usda.gov/countrysummary/Default.aspx?id=IN](https://ipad.fas.usda.gov/countrysummary/Default.aspx?id=IN)