

Molecular mapping of quantitative trait loci for stigma exertion trait in rice (*Oryza sativa* L.)

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Abstract

Stigma exertion is the most important floral trait of female parent/cytoplasmic male sterile (CMS) line for out crossing which increases hybrid seed production at farmer's field. The present study was aimed to map the genomic regions controlling the stigma exertion in F₂ mapping population derived from IR68897B, a recipient maintainer line of IR68897A with low stigma exertion (36.78 %) and BF-16B, a donor parent for high stigma exertion (80.25%). Phenotyping and genotyping of stigma exertion trait was done for 188 individuals of F₂ population developed from IR68897B X BF16B. Frequency distribution of F₂ population showed continuous variation which indicated polygenic nature of inheritance. Phenotypic evaluation revealed that significant variation for stigma exertion trait and transgressive segregation was observed. A total of 15 QTLs were identified for total stigma exertion rate (TSE) which includes single and dual stigma exertion distributed on chr 2, 3, 4, 5, 6, 8, 11 and 12 through QTL mapping. The phenotypic variance and LOD value explained by each QTL ranged from 1.19 to 5.83% and 2.54 to 9.29 respectively. In the present study, total 15 QTLs were minor effect QTLs (<10% phenotypic variance), indicating that stigma exertion is controlled by many small effect QTLs. The QTL *qTSE11-1* on chr 11 (9.29) and *qTSE3-1* on chr 3 (6.83) were identified with higher LOD values and *qTSE3-2* on chr 3 and *qTSE6-2* on chr 6 explained 5.83 and 5.76 % of phenotypic variance, respectively

Keywords: QTL Mapping, stigma exertion, maintainer line, phenotyping, grain yield

Introduction

Rice (*Oryza sativa* L.) is the second most important cereal crop and occupies a predominant position among major food grains. It is one of the crops responsible for the green revolution in the 1960s and 1970s. Global rice demand estimated to rise from 759.6 million tons in 2018 (FAO RMM, 2018) to 852 million tons in 2035 (Kush, 2013). This demand can be achieved by concentrating breeding efforts to improve rice yield potential with the shrinking natural resource base. With an ever-increasing population,

crop scientists continuously work to meet the demands. Different strategies have been suggested to increase the yield potential of rice (Kush, 2005 and 2013). Among various strategies hybrid breeding is one of the strategies to improve rice productivity. Rice hybrids offer an yield advantage of 10-15 % over inbred varieties. Unfortunately, this technology was less adopted by the farming community. The high cost of hybrid seed is one of the major reasons affecting large scale adoption of the technology. This is mainly because of low hybrid seed yields i.e 1-2 t/ha. It is



difficult to reliably produce an acceptable quantity of hybrid rice seeds owing to its self-pollinating nature (Azzini and Rutger, 1982 and Kato and Nimai, 1987). The F_1 seed yield depends on outcrossing potential of female parent of a hybrid. Hybrid rice research should focus on to improve the F_1 seed yields by improving the out crossing potential of CMS lines and by managing seed production practices. This would maintain the hybrid seed cost. Therefore, improvement of hybrid seed production efficiency is an essential factor for large scale commercialization of hybrid rice (Tiwari *et al.*, 2011).

In hybrid rice breeding programs, morphological characteristics of florets are of utmost importance in increasing the out crossing rate in hybrid rice seed production. The floral traits which can improve outcrossing potential of CMS lines include days to heading, pollen load, pollen longevity and morphological traits of floret, *viz.*, size of stigma and style, stigmatic receptivity, stigma exertion, angle of floret opening and duration (Virmani, 1994). Among them, stigma exertion is emphasized as a significant component in increasing pollination and seed set (Kato and Namai, 1987; Virmani, 1994; Sheeba *et al.*, 2006). Stigma receptivity varies from 3-7 days, increasing the chance for pollination (Yan and Li, 1987; Xu and Shen, 1988; Li *et al.*, 2004). Yang, (1997) reported that with 1 percent increase in stigma exertion rate in male sterile lines, hybrid seed yields would increase by 47–68 kg /ha. Therefore stigma exertion rate is extremely important floral trait for improving seed yields in hybrid seed production plots.

Stigma exertion includes single, dual stigmas and other stigma traits that play important roles in hybrid seed production and received consistent attention from rice researchers (Virmani and Athwal, 1973; Virmani, 1994; Yan and Li, 1987; Tian, 1993; Li *et al.*, 2001; Uga *et al.*, 2003; Xu 2003; Yamamoto *et al.*, 2003; Miyata *et al.*, 2007; Sidharthan *et al.*, 2007; Yan *et al.*, 2009, Huang *et al.*, 2012; Dang *et al.*, 2016; Zhou *et al.*, 2017; Guo *et al.*, 2017; Li *et al.*, 2014; Liu *et al.*, 2015; Sruthi *et al.*, 2016; Rahman *et al.*, 2017;

Zhang *et al.*, 2018; Liu *et al.*, 2019, Bakti and Tanaka, 2019; Xu *et al.*, 2019). Earlier studies have suggested that stigma exertion is partially dominant (Virmani and Athwal, 1974) or completely dominant (Li *et al.*, 1997b), and is a qualitative character controlled by a major gene (Hassan *et al.*, 1984). Many studies reported that stigma exertion is a quantitative trait controlled by polygenes (Li *et al.*, 1995; Virmani and Athwal, 1974; Li and Chen, 1985; Wang *et al.*, 2008; Xu and Shen, 1987; Sruthi *et al.*, 2014). The small size of rice spikelets and the large effect of the environment on flowering in rice add to the difficulty in traditional selection-based breeding. As a result, the development of DNA markers associated with desirable floral traits for breeding programs has received increased attention at the national and international levels. Recently many studies have reported QTLs for stigma exertion traits. Tan *et al.*, (2020) mapped 7 QTLs on 5 chrms. (1, 3, 5, 9 and 10) using single segment substitution lines (SSSLs) derived from *O. glumaepatula*. Xu *et al.*, (2019), identified a major QTL for stigma exertion rate *viz.*, qSER-3.1 on chrms 3 in a 3.9 Mb region. Zhang *et al.*, (2018), mapped qSE7 between RM5436 and RM5499 markers, within a physical distance of 1000-kb, with the use of INDEL markers it was finally mapped to a region of 322.9-kb between InDel SER4-1 and RM5436. Rahman *et al.*, (2017) dissected a major QTL qSE11 on chrms 11 to a region of 350.7-kb. Liu *et al.*, (2019) fine mapped qSER7 on chrms 7 and narrowed it down to a region of 28.4-kb between the markers RM3859 and Indel4373. Rahman *et al.*, (2016) identified 8 QTLs (qSES6, qSSE11, qDSE1a, qDSE1b, qDSE10, qDSE11, qTSE1 and qTSE11) for single, dual and total stigma exertion. Bakti and Tanaka, (2019) mapped QTLs for stigma exertion ratio on chrms 2, 3, 4, 8, and 11 from *Oryza rufipogon* accession 'W0120'. Although many QTLs have been identified for stigma exertion, their expression may vary one genetic back ground to other. We have attempted to map QTLs in the back ground of popularly used maintainer lines in hybrid breeding programme in India. Hence, the present investigation was carried out with an objective to identify the QTLs for stigma exertion traits.

Materials and Methods

Plant material

The F₂ mapping population consisting of 188 lines was developed from a cross between IR68897B as a recipient parent with low stigma exertion and BF-16B as a donor parent with high stigma exertion (**Figure 2**). IR68897B is an early duration maintainer line for wild abortive male-sterile cytoplasm with long slender grain type developed by IRRI, Philippines and BF-16B is also a maintainer line for WA-CMS improved for stigma exertion trait by Barwale Company. Hybrid nature of F₁s was confirmed using reported markers (**Table 1**) which are polymorphic between parents and true F₁s were raised in the field to develop F₂ population for QTL mapping.

Field experiments

A total of 188 F₂ mapping population along with its parents was grown in field at Research Farm, Indian Council of Agriculture Research (ICAR)- Indian Institute of Rice Research (IIRR), Hyderabad during *kharif* 2014 for phenotypic evaluation of stigma exertion trait.

Phenotyping of stigma exertion trait

For phenotypic evaluation of stigma exertion trait, panicles were collected at post anthesis. Immediately after collecting panicles, panicles were wrapped in water soaked blotting paper and stored at -20°C. This is to avoid damage to stigma and to keep panicles afresh. For assessing the type of stigma exertion in each spikelet of a panicle by the whole panicle method, all the individual spikelets in each panicle were separated and observed under illuminated magnifier lens to categorize them into dual (DStgE), single (SStgE) (**Figure 1**) or no stigma exertion (NStgE) types. Then, these counts were converted to:

$$\text{SStgE (\%)} = [\text{SStgE} / (\text{SStgE} + \text{DStgE} + \text{NStgE})] \times 100$$

$$\text{DStgE (\%)} = [\text{DStgE} / (\text{SStgE} + \text{DStgE} + \text{NStgE})] \times 100$$

$$\text{Total stigma exertion [TStgE] (\%)} = \text{SStgE (\%)} + \text{DStgE (\%)} \\ \text{and NStgE (\%)} = 100 - \text{TStgE (\%)}$$

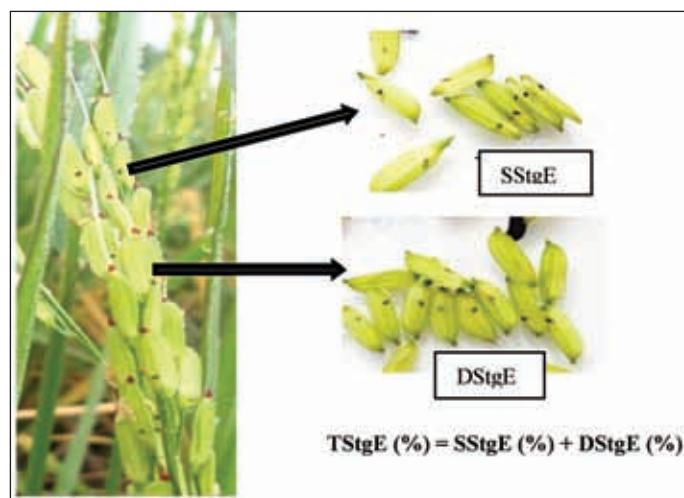


Figure 1: Figure showing the phenotyping of stigma exertion trait

Genotyping

Genomic DNA was isolated from total 188 F₂ population and the parents using CTAB method of Zheng *et al.*, (1991) with minor modifications. Parental polymorphism survey was done using 513 SSR markers, which are distributed across 12 chr. Out of 513 SSR markers only 124 markers showed (**Table 3**) polymorphism between parents. PCR amplification was carried out using thermal cycler (Veriti Thermo Cycler, Applied Bio systems). Master mix was prepared for 10 µl reaction volume containing 2 µl DNA template, 4.5 µl of nuclease free water, 1.0 µl 10X buffer, 1 µl dNTPs, 1 µl for both forward and reverse primers and 0.5 µl of 2U/ µl Taq DNA polymerase. PCR thermal profile is as followed: initial denaturation cycle of 95°C for 5 min was followed by 35 cycles at 95°C for 30 sec, 55°C for 30 sec, 1 min of extension at 72°C with an additional step of 10 min at 72°C. PCR products were resolved in 3% agarose gel in 0.5X TBE buffer stained with Ethidium Bromide and documented using gel documentation system (Bio-Rad, USA).

Linkage map construction and QTL mapping

The polymorphic 56 SSR (**Table 2**) markers were used to analyze the genomic DNA of the parents, IR68897B and BF-16B and F₂ population (188 plants). Each gel was scored for maternal (A), paternal (B) and heterozygous (H) banding pattern and later these converted into 2 (A), 0 (B) and 1 (H) score. QTL and

their effects were obtained using framework linkage map constructed for the genotypic data of 188 F₂ mapping population using 56 polymorphic markers and phenotypic data of 188 F₂ population using QTL ICI (Integrated Composite Interval Mapping) Mapping V4.2 (Meng *et al.*, 2015) employing Inclusive Composite Interval Mapping (ICIM) method. The LOD profiles were used to identify most of the likely position for a QTL in relation to the linkage map, which was the position where the highest LOD value was obtained. For QTL mapping, logarithm of the odds (LOD) threshold value was set to 2.5 using 1000 permutations at 0.05 level of significance.

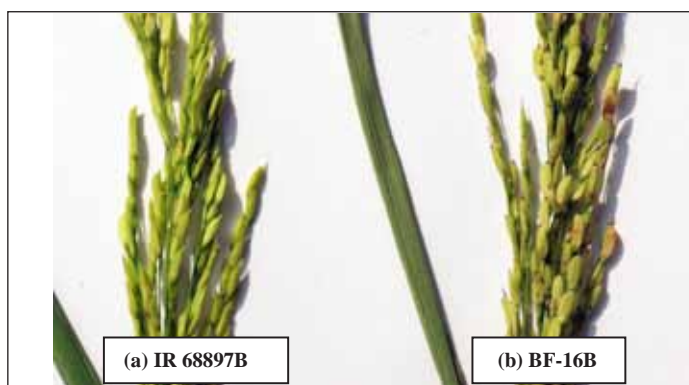


Figure 2: Figure showing the panicle type of parents; (a) Recipient (IR 68897B) and (b) donor parent (BF-16)

Results and discussion

Phenotypic performance of stigma exertion in the F₂ mapping population

The two parents used in the cross had contrasting stigma exertion rates, *viz.*, 36.78 and 80.25 %, while the F₁ was lower than better parent and higher than recipient parent *i.e.*, intermediate with 60.44 % stigma exertion rate.

Frequency distribution of stigma exertion in F₂ population

The frequency distribution of the stigma exertion rate in the F₂ population showed a continuous variation. The total phenotypic data were divided as per the scale of IRRI. The scale of stigma exertion plotted against X-axis and number of plants on Y-axis. The graph showed a bell-shaped normal distribution curve and skewed towards low stigma exertion. The frequency distribution of stigma exertion (**Figure 3**) indicated

that both parents have several chromosomal regions increasing the frequency of stigma exertion and the trait is affected by the environment. In the present cross transgressive segregants were observed in the F₂ population.

Genotyping

Of the 513 SSR markers screened for parental polymorphism between IR 68897B and BF-16B, only 124 (24.17 %) showed clear polymorphism. A set of 56 polymorphic SSR markers, evenly distributed on rice genome covering all 12 chr and showing clear polymorphism between stigma exertion recipient and donor parents were used for genotyping of 188 F₂ population.

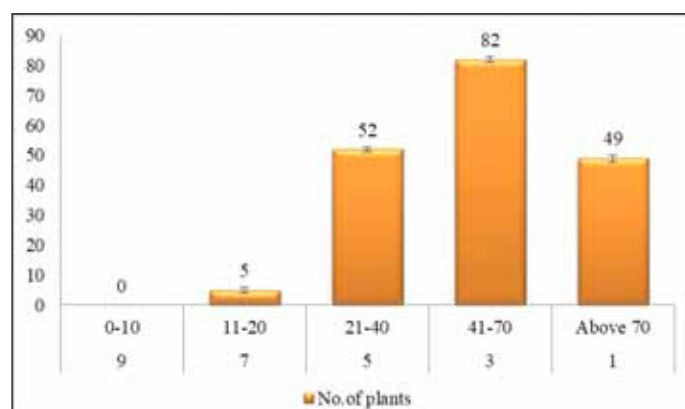


Figure 3: The graph depicts the frequency distribution of stigma exertion trait in 188 F₂ mapping population; 1, 3, 5, 7 and 9 the scale given by IRRI for phenotyping of stigma exertion trait.

QTL mapping of stigma exertion in F₂ mapping population

A total of 15 QTLs were identified for total stigma exertion rate which included single, dual stigma exertion using 56 polymorphic SSR markers with 188 F₂ mapping population and they were distributed on all 12 chr except on chr 1, 7, 9 and 10. The phenotypic variance and LOD value explained by each QTL ranged from 1.19 to 5.83% and 2.54 to 9.29, respectively. The majority of QTLs were contributed from the donor parent. On chr 2, three QTLs were identified. The QTL, *qTSE2-1* was mapped at 24 cM and flanked by RM151 and RM12398 which accounts for 4.5% of the phenotypic variance with a LOD value of 3.75, the *qTSE2-2* was mapped at 129 cM, flanked

by markers RM13131 and RM208 and showed 4.94% of the phenotypic variance with a LOD value of 3.30 and the third QTL on chr2, *qTSE2-3* was flanked by RM208 and RM13238 at 218 cM map position and explained 4.98% of the phenotypic variance with 4.69 LOD value.

The QTL cluster *qTSE-3-1*, *qTSE-3-2* and *qTSE-3-3* on chr3 were flanked by RM1350-RM15466, RM1350-RM15466 and RM15466-RM7000 respectively and collectively explained 16.12% of the phenotypic variance with LOD value 6.83, 5.66 and 4.66 respectively.

A single QTL, *qTSE-4-1* was identified on chr4 at map position of 47 cM flanked by RM16338 and HRM17405 and explained 4.84 % of phenotypic variance with a LOD score of 3.5. On chr5, two QTLs, *qTSE-5-1* and *qTSE-5-2* were identified at map positions of 117 and 184 cM flanked by the markers RM5592- HRM18222 and HRM18222-RM17950. Two QTLs *qTSE-5-1* and *qTSE-5-2* on chr5 collectively accounted for 6.51 % of phenotypic variance with LOD scores of 2.54 and 2.59, respectively.

Two QTLs *qTSE-6-1* and *qTSE-6-2* were identified on chr6 flanked by RM 20615-RM19569 with LOD value of 5.16 and 6.30 and explaining phenotypic variance of 1.19 and 5.76 %, respectively. Two QTLs *qTSE-8-1* and *qTSE-8-2* on chr8 flanked by the markers RM22977-RM23643 and RM5493-RM6925 with LOD values of 3.88 and 3.77 and explaining 5 and 1.43 % of phenotypic variance, respectively. A single QTL *qTSE-11-1* was detected on chr11 flanked by RM27183-RM26213 and explained 3.10 % phenotypic variance with LOD value of 9.29. A single QTL *qTSE-12-1* on chr12 flanked by RM7102-RM28157 and explained 4.28% phenotypic variance with LOD value of 2.72. The study detected contributing alleles for the trait were distributed in both i.e., donor and recipient parents. Eleven QTLs distributed on chr3, 4, 5, 6, 8, 11 and 12 from donor parent (BF-16B) and four QTLs present on chr2 and 8 were from recipient parent (IR 68897B). The QTL clusters identified on chr2(3 QTLs), 3(3 QTLs), 5(2 QTLs), 6(2 QTLs), 8(2 QTLs) collectively explained 14.42%, 16.12%, 6.51%, 6.95%, and 6.43%, respectively. The details of the putative QTLs for stigma exsertion were given in the **Table 4 and Figure 4**.

Table 4. Putative QTLs for total stigma exsertion detected in the F₂ population derived from IR 68897B and BF-16B

S.No.	QTL	Chr.	Position (cM)	Left Marker	Right Marker	LOD	PVE (%)	Add	Dom
1	qTSE-2-1	2	24	RM151	RM12398	3.75	4.50	0.87	21.17
2	qTSE-2-2	2	129	RM13131	RM208	3.30	4.94	1.31	21.46
3	qTSE-2-3	2	218	RM208	RM13238	4.69	4.98	4.23	-20.08
4	qTSE-3-1	3	28	RM1350	RM15466	6.83	5.19	-13.84	6.05
5	qTSE-3-2	3	94	RM1350	RM15466	5.66	5.83	-3.91	22.96
6	qTSE-3-3	3	153	RM15466	RM7000	4.66	5.10	-2.23	21.92
7	qTSE-4-1	4	47	RM16338	HRM17405	3.50	4.84	-3.86	-21.48
8	qTSE-5-1	5	117	RM5592	HRM18222	2.54	1.79	-7.98	-6.05
9	qTSE-5-2	5	184	HRM18222	RM17950	2.59	4.72	-0.35	21.37
10	qTSE-6-1	6	0	RM20615	RM19569	5.16	1.19	-1.02	-10.42
11	qTSE-6-2	6	58	RM20615	RM19569	6.30	5.76	-2.12	23.28
12	qTSE-8-1	8	27	RM22977	RM23643	3.88	5.00	-1.79	22.06
13	qTSE-8-2	8	155	RM5493	RM6925	3.77	1.43	0.16	12.45
14	qTSE-11-1	11	96	RM27183	RM26213	9.29	3.10	-10.42	3.97
15	qTSE-12-1	12	35	RM7102	RM28157	2.72	4.28	-1.10	20.05

PVE- Phenotypic variance explained by each QTL, LOD-Logarithm of odds ratio, qTSE-QTL for total stigma exsertion. Add-the additive effect of each QTL and dominant effect of each QTL

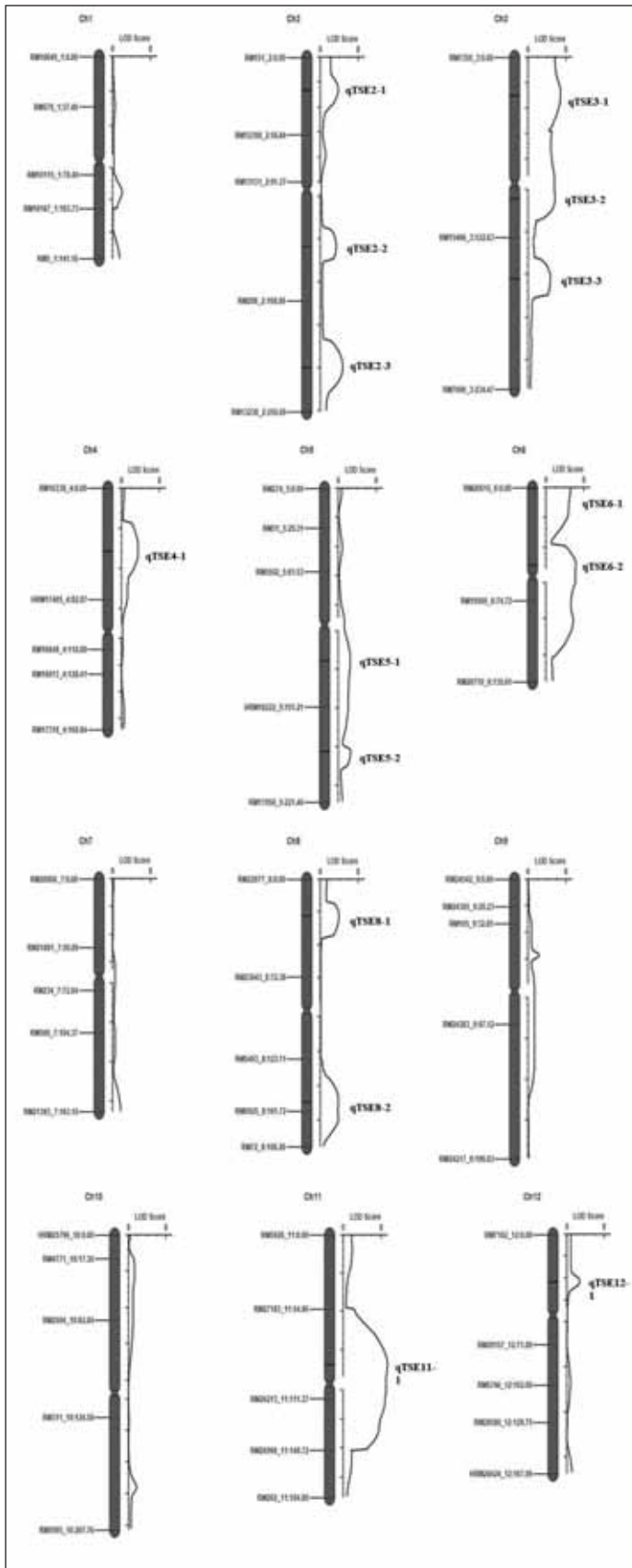


Figure 4: Distribution of QTLs for stigma exertion trait on the linkage map

Breeding for hybrid rice is one of the pragmatic solutions for addressing the problem of food shortage, caused by marked increase in - global population with decreasing trend in available resources, land and water for agriculture. Breeding approaches may solve this problem by increasing yield per unit area through hybrid system of breeding. But the large scale adoption of hybrid rice has been hampered by low F_1 seed set and the consequent high price of hybrid seed. To overcome the extremely low seed set percentage of CMS line, there is a need to improve the parental lines for flowering behavior and floral traits and modifications in seed production practices are useful for pushing seed yields beyond 2.5 tonnes per hectare and also bring down the F_1 seed cost.

In our study, transgressive segregants were observed in both the directions signifying complementary action of the genes with additive effect that were dispersed in both the parents and a normal distribution for the phenotype was observed. According to Singh and Narayanan (1993) transgressive segregations are only possible from the crosses between two parents with mean values for a quantitative trait. Such segregants are not possible in case of qualitative traits. De Vicente and Tanksley, (1993) studied that the transgressive segregation is commonly observed in the population segregating for quantitative trait. Whereas Ricks and Smith (1953), mentioned that transgressive segregation in certain progeny is because of accumulation of complementary alleles at multiple loci inherited from the two parents.

These results were in conformity with the results of Virmani and Athwal, (1974), Li and Chen, (1985), Wang *et al.*, (2008), Ling *et al.*, (1989), Miyata *et al.*, (2007), Lou *et al.*, (2014), Li *et al.*, (2014) and Sruthi *et al.*, (2014) who observed wide continuous variation with normal distribution and concluded that the stigma exertion trait of rice is a quantitative trait, controlled by polygenes. Similarly, in tomato, stigma exertion has been reported to be a quantitative trait and controlled by a few genes (Rick and Dempsey, 1969; Scott and George, 1978; Levin *et al.*, 1994). However, earlier reports on the genetics of stigma exertion in rice showed contradiction to the present finding. For instance, Hassan and Siddiq, (1984)

reported that fully exerted stigma to be monogenic and dominant over partially exerted stigma. Xu and Shen, (1987) also studied the inheritance of the stigma exertion trait and reported that it was a dominant trait and in their study F_2 population showed continuous variation for stigma exertion, this may be due to some minor genes supplementing the major gene in the expression of the trait. Experimental evidence suggested that environmental effects would also produce a continuous variation even if the number of genes governing a character was very small or even one.

We constructed 188 lines of the F_2 mapping population using IR68897B x BF-16B cross and genotyped these 188 lines with 56 polymorphic molecular markers to identify the QTLs for the trait of interest (stigma exertion). Results showed that 15 QTLs were associated with stigma exertion traits and were located on chr2, 3, 4, 5, 6, 8, 11 and 12. The phenotypic variance and LOD value explained by each QTL ranged from 1.19 to 5.83% and 2.54 to 9.29 respectively. The present QTL analysis results for stigma exertion were in accordance with the result of Tan *et al.*, (2020). They mapped the major QTLs for stigma exertion rate from *Oryza glumaepatula* on chr1 and 3. Xu *et al.*, (2019) identified a major QTL on chr3 using double haploid population. Lou *et al.*, (2014) identified a total of 5 QTLs and 9 epistatic QTL pairs were found to associate with the stigma trait in F_2 population of Huhan 1b and K17B. Li *et al.*, (2001) reported QTL on Chr5 and 8 using back cross-population of Dongxiang and Guichao 2. Li *et al.*, (2001) identified QTLs on chr2 and 3 using DH population of Zaiyeqing 8 and Jingxi 17. Uga *et al.*, (2003) identified QTL for the rate of stigma exertion on chr5 and 10 using RILs of Pei-Kuh and a wild rice W1944 and Deng *et al.*, (2011) observed on chr1, 2, 3 and 5 using F_2 population of (CMS) maintainer line II-32B and G46B. Miyata *et al.*, (2007) reported QTLs for stigma exertion on chr3 using F_2 population of Koshihikari and IR24 and Yue *et al.*, (2009) demonstrated on chr3, 4, 7 and 9 using F_2 population of CMS maintainer line Huhan 1B and II-32B. Wang *et al.*, (2008) studied high SE in chr2, 5 and 8 by introducing the genes of high stigma exertion rate from *indica* into *japonica*.

The National plant biology symposium proceeding in China (2011) confirmed the QTLs for PES (percentage of exerted stigma) to be present on all chr except on chr7. The studies on stigma exertion by Li *et al.*, (2014) reported the presence of the trait in chr3, 6, 7, 9, 10 and 12 using RIL population of ZX and CX29B. Lou *et al.*, (2014) also reported the presence of PSE in chr5 and 6, 7 using F_2 population of CMS lines, Huhan1B and K17B, which are similar to the results of the present study. Li *et al.*, (2014) resolved exerted stigma determined by a few main and epistatic pairs QTLs, indicating mainly influenced by minor QTLs in three-way cross/backcross population of Zhenshan 97B and 9311. In Rahman *et al.*, (2016) studies showed a total of two QTLs for TSE located on chr1 and 11 in two environments and QTL on chr11 is novel to date for exerted stigma in our result also.

In addition, results detected contributing alleles for trait were distributed in both i.e., donor and recipient parents. Eleven QTLs distributed on chr3, 4, 5, 6, 8, 11 and 12 from BF-16B and four are on chr2 and 8 from IR 68897B. Previous studies (Rahman *et al.*, 2016; Li *et al.*, 2014; Yamamoto *et al.*, 2003; Yu *et al.*, 2006) reported that favorable alleles present and contribute to trait expression by both the parents. Hence, for improvement of exerted stigma trait it is very much essential to select suitable breeding program for pyramiding of traits in parental lines from different sources.

In the present study one QTL cluster has been identified on chr3 flanked between RM 1350 and RM 7000 for TSE, it collectively explaining 16.12 % phenotypic variance. Earlier also reported by Lou *et al.*, (2014) for PDES on chr5 and by Yu *et al.*, (2006), mapped the region by Li *et al.*, (2001) and Uga *et al.*, (2003b). Li *et al.*, (2014) also detected two QTLs clusters on chr1 and 6 for extruded stigma.

Many QTL identified by different researchers for stigma exertion rate (SER), Percentage of exerted stigma (PES), Percentage of single exerted stigma (PSES) and Percentage of double exerted stigma (PDES) in rice. In the present investigation, 15 QTLs were identified, of which 11 were contributed from



the donor parent (BF-16B) and 4 from the recipient parent (IR 68897B) for stigma exertion to the progeny. Both the parents contributed towards the stigma exertion. However, more number of QTLs was contributed by the donor parent to the progeny for total stigma exertion (TSE) in *indica* rice. All the 15 QTLs identified for total stigma exertion in this study were minor affect QTLs accounting for less than 10 % of the phenotypic variance. This clearly indicates the polygenic nature of the trait controlled by several genomic regions with small effects. Stigma exertion is highly influenced by environment which might be affecting the QTL expression. In general, QTLs with large effects are more stable across environments than QTLs with smaller effects. stigma exertion is controlled by many genes and has low heritability. Hence, it is advisable to phenotype the trait in multiple environments in order to identify the stable QTLs. In the present study, the markers flanked by the QTLs can be utilized to screen the uncharacterized germplasm for stigma exertion trait to identify new donors and can be used in marker assisted breeding programmes to improve maintainer lines. These maintainers can be converted into new CMS lines with improved stigma exertion rate to increase hybrid seed production efficiency. The QTLs identified in the present study need validation before they can be utilized in Marker-Assisted breeding programs for improvement of the maternal parent in hybrid rice technology. This may exertion help in overcoming the pollination barriers like time (asynchronous flowering) and will allow accumulation of more pollen on stigmatic surface of female parent improving the hybrid seed production efficiency.

Authors Contribution

Conceptualization of research (KKB, SMM & ASH): Designing of the experiments (KKB, SMM & ASH): Contribution of experimental material (Barwale Foundation): Execution of field /lab experiments and data collection (GS & JK): Analysis of data and interpretation (GS, JK, KKB, SMM & SK): Preparation of the manuscript and correction (GS, JK, KKB, SMM, SK, VS, SJ, AKS, PK, LS, SP, GC, AMS, RMS & ASH).

Compliance with Ethical Standards: The authors declare the research work is in Compliance with Ethical Standards

Conflict of Interest: The authors declare they have no conflicts of interest.

Ethical approval: This article does not contain any studies with human participants or animals performed by any of the authors.

References

- Azzini LE and Rutger JN. 1982. Amount of outcrossing on different male sterile of rice. *Crop Science*, 22: 905-907
- Bakti C and Tanaka J. 2019. Detection of dominant QTLs for stigma exertion ratio in rice derived from *Oryza rufipogon* accession 'W0120'. *Breeding Science*, 69(1): 143-150.
- Cheng SH, Zhuang JY, Fan YY, Du JH and Cao LY. 2007. Progress in research and development on hybrid rice: a super domesticate in China. *Annal of Botany*, 100 (5): 959-966
- Dang XJ, Liu EB, Liang YF, Liu QM, Breria CM, Hong DL. 2016. QTL detection and elite alleles mining for stigma traits in *Oryza sativa* by association mapping. *Frontiers in Plant Science*, 7: 1188. doi: 10.3389/fpls.2016.01188.
- De Vicenta MC and Tanksley SD. 1993. QTL analysis of transgressive segregation in an interspecific tomato cross. *Genetics*, 134:585-596
- Deng YD, Xiao CL, Deng HB, Zhang HQ, Deng XJ and Liu YL. 2011. Detection of QTL related to stigma exertion rate (SER) in rice (*Oryza sativa* L.) by bulked segregant analysis. *Research of Agricultural Modernization*, 32(2): 230-233.
- Food and Agricultural Organization (FAO). 2018. *Rice Market Monitor*, 21(1): 1-35.
- Guo L, Qiu FL, Gandhi H, Kadaru S, De Asis EJ, Zhuang JY, Xie FM. 2017. Genome-wide association study of outcrossing in cytoplasmic male sterile lines of rice. *Scientific Reports*, 7:3223. doi: 10.1038/s41598-017-03358-9.

- Hassan MA and Siddiq EA. 1984. Inheritance of anther size and stigma exertion in rice (*Oryza sativa* L.). *Indian Journal of Genetics and Plant Breeding*, 44(3): 544-547
- Huang X, Kurata N, Wang ZX, Wang A, Zhao Q, Zhao Y, Liu K, Lu H, Li W, Guo Y and Lu Y .2012. A map of rice genome variation reveals the origin of cultivated rice. *Nature*, 490(7421): 497-501.
- K Sruthi, KB Eswari, KB. Kemparaju, K Jayaramulu and M Sheshu Madhav. 2016. Identification of SSR marker linked to the stigma exertion in maintainer lines of hybrid rice. *Ecology Environment and Conservation*, S133-S139. Copyright@ EM International. ISSN 0971-765X.
- Kato H and Namai H. 1987. Floral characteristics and environmental factors for increasing natural outcrossing rate for F1 hybrid seed production of rice *Oryza sativa* L. *Japanese Journal of Breeding*, 37: 318-330
- Khush GS. 2013. Strategies for increasing the yield potential of cereals: case of rice as an example. *Plant Breeding*, 132(5): 433-436.
- Khush GS. 2005. What it will take to feed 5.0 billion rice consumers in 2030. *Plant Molecular Biology*, 591-6. 10.1007/s11103-005-2159-5.
- Levin I, Cahaner A, Rabinowitch HD and Elkind Y. 1994. Effects of the *ms10* gene, polygenes and their interaction on pistil and anther-cone lengths in tomato flowers. *Heredity*, 73: 72-77.
- Li PB, Feng FC, Zhang QL, Chao Y, Gao GJ, He YQ. 2014. Genetic mapping and validation of quantitative trait loci for stigma exertion rate in rice. *Molecular Breeding*, 34:2131-2138. doi: 10.1007/s11032-014-0168-2.
- Li Z, Chen G, Wang Z. 2004. Study on the target characters of almost normal outcrossing sterile line of japonica hybrid rice. *Reclamation Cultivation Rice*, 3:7-10
- Li C, Sun CQ, Mu P, Chen L and Wang XK. 2001. QTL analysis of anther length and ratio of stigma exertion, two key traits of classification for cultivated rice (*Oryza sativa* L.) and common wild rice (*O. rufipogon* Griff.). *Acta Genetica Sinica*, 28(8): 746-751
- Li H, Ribaut JM, Li Z and Wang J. 2008. Inclusive composite interval mapping (ICIM) for digenic epistasis of quantitative traits in biparental populations. *Theoretical and Applied Genetics*, 116: 243-260.
- Li T and Chen YW. 1985. Genetics of stigma exertion in rice. *Rice Genetics Newsletter*, 2: 84-85.
- Li ZK, Pinson SR, M Stansel JW and Park WD. 1995. Identification of quantitative trait loci (QTL) for heading date and plant height in cultivated rice (*Oryza sativa* L.). *Theoretical and Applied Genetics*, 91: 374-381
- Li ZK, Pinson SR, M Stansel JW and Park WD. 1997b. Genetics of hybrid sterility and hybrid breakdown in an inter-sub specific rice (*Oryza sativa* L.) population. *Genetics*, 145: 1139-1148
- Ling ZM. 1989. Study on stigma exertion after anthesis in rice. II. The inheritance of stigma exertion in some rice crosses. *Acta agriculturae Universitatis Pekinensis*, 15(1): 23-27.
- Liu QM, Qin JC, Li TW, Liu EB, Fan DJ, Edzesi WM, Liu JH, Jiang JH, Liu XL, Xiao LJ, Liu LL, Hong DL. 2015. Fine mapping and candidate gene analysis of qSTL3, a stigma length-conditioning locus in rice (*Oryza sativa* L.). *Plos One*, 10: e0127938. doi: 10.1371/journal.pone.0127938.
- Liu Y, Zhang A, Wang F, Kong D, Li M, Bi J, Zhang F, Wang J, Luo X, Pan Z and Yu X. 2019. Fine mapping a quantitative trait locus, qSER-7, that controls stigma exertion rate in rice (*Oryza sativa* L.). *Rice*, 12(1):1-10.
- Lou J, Yue GH, Yang WQ, Mei HW, Luo LJ and Lu HJ. 2014. Mapping QTLs influencing stigma in rice. *Bulgarian Journal of Agricultural Science*, 20:1450-1456
- MH Rahman, P Yu, YX Zhang, LP Sun, WX Wu, XH Shen, XD Zhan, DB Chen, LY Cao and SH Cheng. 2016. Quantitative trait loci mapping of



- the stigma rate and spikelet number per panicle in rice (*Oryza sativa* L.). *Genetics and Molecular Biology*, 15 (4): 1-11
- Meng L, Li H, Zhang L and Wang J. 2015. QTL IciMapping: integrated software for genetic linkage map construction and quantitative trait locus mapping in biparental populations. *Crop Journal*, 3(3), 269-283.
- Miyata M, Yamamoto T, Komori T and Nitta N. 2007. Marker-assisted selection and evaluation of the QTL for stigma exertion under japonica rice genetic background. *Theoretical and Applied Genetics*, 114(3): 539-548
- National plant biology symposium proceeding in China (2011)
- Phundan Singh and Narayanan SS .1993. Biometrical Techniques in Plant Breeding (first ed.) Kalyani Publishers, New Delhi, India
- Pingbo Li, Fuchun, F, Qinglu, Z, Yuan, C, Guanjun, G and Yuqing H. 2014. Genetic mapping and validation of quantitative trait loci for stigma exertion rate in rice. *Molecular Breeding*, 34:2131–2138
- Rahman MH, Zhang YX, Sun LP, Zhang KQ, Rahman MS, Wu WX, Zhan XD, Cao LY, Cheng SH. 2017. Genetic mapping of quantitative trait loci for the stigma exertion rate in rice (*Oryza sativa* L.). *Journal of Integrative Agriculture*, 16:60345–60347. doi: 10.1016/S2095-3119(16)61540-X.
- Rick CM and Smith PG. 1953. Novel variation in tomato species hybrids. *The American Naturalist*, 87(837): 359-373.
- Rick CM and Dempsey WH. 1969. Position of the stigma in relation to fruit setting of tomato. *Botanical Gazette*, 130: 180–186.
- Scott JW and George WL. 1978. Breeding and combining ability of heterostylous genotypes for hybrid seed production in *Lycopersicon esculentum* Mill. *Euphytica*, 29: 135–144.
- Sheeba A, Vivekanandan P and Ibrahim SM. 2006. Genetic variability for floral traits influencing out crossing in the CMS lines of rice. *Indian Journal of Agricultural Research*, 40 (4): 272 – 276
- Sidharthan B, Thiyagarajan K and Manonmani S. 2007. Cytoplasmic male sterile lines for hybrid rice production. *Journal of applied sciences research*, 3(10): 935-937
- Singh P and Narayanum SS. 1993. Biometrical techniques in plant breeding. Kalyani publishers, New Delhi. 23-32.
- Sreedhar S, Reddy TD and Ramesha MS. 2011. Genotype x Environment interaction and stability for yield and its components in hybrid rice cultivars (*Oryza sativa* L.). *International Journal of Plant Breeding and Genetics*, 5: 194208
- Sruthi K, KB Eswari, KB Kemparaju, K Jayaramulu, M Sheshu Madhav, MHV Bhav, Swapna K, DS Steffi Graf, AS Hariprasad, P Senguttuvel and P Revathi. 2014. Genetics of stigma exertion in maintainer lines of hybrid rice. *Progressive Research*, 9 (Conf. Spl.): 1275-1276.
- Tan Q, Zou T, Zheng M, Ni Y, Luan X, Li X, Yang W, Yang Z, Zhu H, Zeng R and Liu G. 2020. Substitution mapping of the major quantitative trait loci controlling stigma exertion rate from *Oryza glumaepatula*. *Rice*, 13(1): 1-10.
- Tang YD, Ying JZ, Shi YY, Xiao S and Zhang H. 2010. Preliminary analysis of percentage of exerted stigma in rice quantitative trait loci. Hunan agricultural university. Natural Science Edition. 04.
- Tian Dacheng. 1993. Studies on mechanism of out-crossing rate in hybrid rice seed production (Sichuan province seed company). *Hybrid rice*. 1
- Tiwari DK, P Pandey, SP Giri and JL Dwivedi. 2011. Heterosis studies for yield and its components in rice hybrids using CMS system. *Asian Journal of Plant Sciences*, 10: 2942
- Uga Y, Fukuta Y, Cai HW, Iwata H, Ohsawa R, Morishima H and Fujimura T. 2003. Mapping QTLs influencing rice floral morphology using recombinant inbred lines derived from a cross

- between *Oryza sativa* L., *Oryza rufipogon* Griff. *Theoretical and Applied Genetics*, 107: 218-226
- Virmani SS and Athwal DS. 1973. Genetic variability of floral characters influencing out crossing in rice. *Crop Science*, 14: 350-353
- Virmani SS and Athwal DS. 1974. Inheritance of floral characteristics influencing outcrossing in rice. *Crop Science*, 14: 350-353.
- Virmani, SS. 1994. Heterosis and hybrid rice breeding. Monographs *Theoretical and Applied Genetics*, 22, Springer-Verlag Berlin.
- Wang YR, Hua ZT, Zhang ZX, Li QY, Li RH, Su YA, Yao JP and Wang ZX. 2008. Breeding and application of japonica male sterile lines with high stigma exertion rate in rice. *Hybrid rice*, 03.
- Xu YB and Shen ZT. 1988. Receptivity of exerted stigma. *International Rice Research Newsletter*, 13(3):7-8
- Xu S, Zheng Y, Liu Y, Guo X, Tan Y, Qian Q, Shu Q and Huang J. 2019. Identification of a major quantitative trait locus and its candidate underlying genetic variation for rice stigma exertion rate. *Crop Journal*, 7(3): 350-359.
- Xu YB and Shen ZT. 1987. Inheritance of stigma exertion in rice. *Rice Genetics Newsletter*, 4:76.
- Xu YB. 2003. Developing marker-assisted selection strategies for breeding hybrid rice. *Plant Breeding Reviews*, 73-174.
- Yamamoto T, Takemori N, Sue N and Nitta N. 2003. QTL analysis of stigma exertion in rice. *Rice Genetics Newsletter*, 20: 33-34.
- Yan WG and Li SF. 1987. Study on out-crossing characteristics among male sterile lines containing same nucleus in rice. *Hybrid Rice*, 4: 8-11
- Yan WG, Li Y, Agrama AH, Luo D, Gao F, Lu, X and Ren G. 2009. Association mapping of stigma and spikelet characteristics in rice (*Oryza sativa* L.). *Molecular Breeding*, 24: 277-292.
- Yang ZP. 1997. Inheritance of photoperiod genic male sterility and breeding of photoperiod sensitive genic male sterile lines in rice (*Oryza Sativa* L.) through anther culture. *Euphytica*, 94:93-99.
- Yu XQ, Mei HW, Luo LJ, Liu GL, Zou GH, Hu SP, Li MS and Wu JH. 2006b. Dissection of additive, epistatic and Q x E interaction of quantitative trait loci influencing stigma exertion under water stress in rice. *Acta Genetica Sinica*, 33(6): 542-550.
- Yue GH, Mei HW, Pan BR, Lou J, Li MS and Luo LJ. 2009. Mapping of QTLs affecting stigma exertion rate of Huhun 1B as a CMS maintainer of upland hybrid rice. *Acta Agriculturae Zhejiangensis*, 21(3): 241-245.
- Zhang K, Zhang Y, Wu W, Zhan X, Anis GB, Rahman MH, Hong Y, Riaz A, Zhu A, Cao Y and Sun L. 2018. qSE7 is a major quantitative trait locus (QTL) influencing stigma exertion rate in rice (*Oryza sativa* L.). *Scientific Reports*, 8(1): 1-11.
- Zheng KL, Shen B and Qian HR. 1991. DNA polymorphism generated by arbitrary primed PCR in rice. *Rice Genetics Newsletter*, 8: 134-136.
- Zhou H, Li PB, Xie WB, Hussain S, Li YB, Xia D, Zhao H, Sun SY, Chen JX, Ye H, Hou J, Zhao D, Gao GJ, Zhang QL, Wang GW, Lian XM, Xiao JH, Yu SB, Li XH, He YQ. 2017. Genome-wide association analyses reveal the genetic basis of stigma Exsertion in Rice. *Molecular Plant*, 10:634-644. doi: 10.1016/j.molp.2017.01.001.